



# Feces consumption by nominally herbivorous fishes in the Caribbean: an underappreciated source of nutrients?

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**Abstract** Parrotfishes and surgeonfishes are major Caribbean herbivores that primarily graze reef algae and thereby play an important functional role in indirectly promoting coral recruitment and growth. Yet, an emerging body of research suggests that these nominal herbivores graze on a diverse array of other food sources and researchers have questioned whether they may target more nutrient-dense foods growing within or upon algae, such as cyanobacteria. In this study, we investigated the species-specific foraging rates of parrotfishes and surgeonfishes on Brown Chromis (*Chromis multilineata*) fecal pellets compared to other major dietary items. We found that almost 85% of observed fecal pellets were ingested by fishes and that over 90% of ingested fecal pellets were consumed by parrotfishes and surgeonfishes alone. While there were species-specific differences in the levels of feces consumption (coprophagy), we found that all three surgeonfishes (*Acanthurus chirurgus*, *A. coeruleus*, and *A. tractus*)

and six of the nine of parrotfish species surveyed (*Scarus coeruleus*, *S. iseri*, *S. taeniopterus*, *S. vetula*, *Sparisoma aurofrenatum*, and *S. viride*) consumed *C. multilineata* feces. To better understand the nutritional value of this behavior, we analyzed the composition of proteins, lipids, carbohydrates, total calories, and micronutrients in *C. multilineata* fecal pellets and compared these to published values for other food sources targeted by these fishes. Our findings suggest that these fecal pellets may have higher values of proteins, carbohydrates, total calories, and important micronutrients, such as phosphorus, compared to various macroalgae and the epilithic algae matrix, though comparable or lower values compared to cyanobacteria. To our knowledge, this is the first study to document coprophagy by tropical herbivorous fishes in the Caribbean region. This research advances our understanding of the foraging ecology of nominally herbivorous fishes and highlights the importance of fish feces as a nutritional resource on coral reefs.

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Hannah S. Rempel and Abigail K. Siebert have contributed equally.

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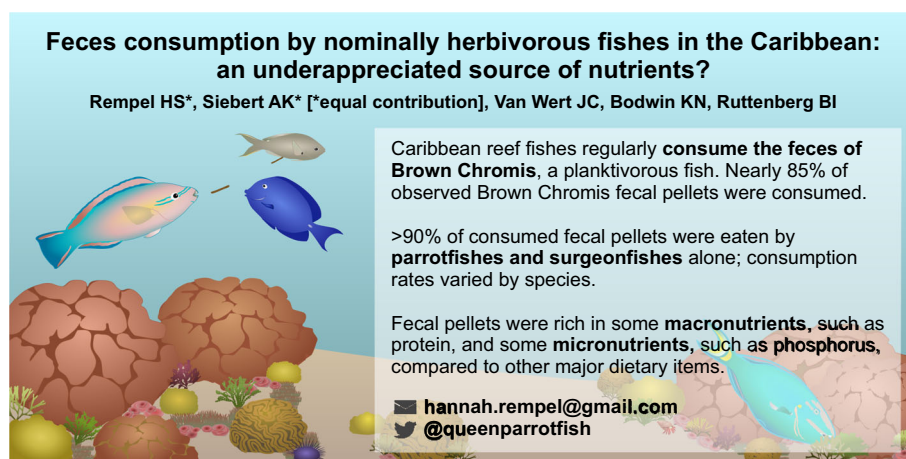
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## Graphical abstract



**Keywords** Coprophagy · Fish feces · Nutritional ecology · Parrotfish · Surgeonfish · Coral reef

## Introduction

Herbivorous fishes play an important role in coral reef ecosystems by grazing algae that can otherwise outcompete corals, thereby indirectly promoting the recruitment, growth, and survivorship of reef-building corals (Hughes et al. 2007; Bonaldo et al. 2014; Adam et al. 2015). Parrotfishes (tribe Scarini within the *Labridae*) and surgeonfishes (*Acanthuridae*) are key herbivores that are abundant on many tropical reefs and predominantly graze upon algae (Lewis and Wainwright 1985; Bellwood et al. 2004; Adam et al. 2015). Many studies have focused on quantifying the grazing dynamics and nutritional ecology of parrotfishes and surgeonfishes to better understand the patterns and drivers of grazing by these critically important fishes. Parrotfishes and surgeonfishes can vary in their grazing behavior interspecifically, ontogenetically, and based on context-dependent factors, such as food and nutrient availability (Burkpile and Hay 2011; Bonaldo et al. 2014; Smith et al. 2018; Duran et al. 2019; Ruttenberg et al. 2019). In addition to algae, nominally herbivorous parrotfishes graze on a diverse array of other food sources such as cyanobacteria, sponges, and corals (Bruggemann et al. 1994a; Burkpile et al. 2019; Nicholson and Clements 2020; Rempel et al. 2020). Surgeonfishes have also been documented grazing on benthic cyanobacterial mats and invertebrates, such as sponges (Cissell et al. 2019; Duran et al. 2019).

Turf algae and the epilithic algal matrix (EAM), which represent a large portion of the diet of parrotfishes and surgeonfishes, also contain notable amounts of

cyanobacteria, feces, microbes, and detritus (Lewis and Wainwright 1985; Wilson 2002; Mumby 2006; Bonaldo et al. 2014; Nicholson and Clements 2020). Therefore, researchers have questioned whether some nominally herbivorous fishes may be detritivores and microphages that target other food sources growing upon algae or within algae-covered substrates that may be more nutrient dense, such as cyanobacteria (Choat et al. 2001; Crossman et al. 2005; Clements et al. 2016; Nicholson and Clements 2020). In addition, one study documented parrotfishes and surgeonfishes eating the feces of other fishes (coprophagy) on Indo-Pacific reefs (Robertson 1982), though this coprophagous behavior has not been documented in other tropical regions. Another study found that a surgeonfish and other fishes consume the feces and vomit of a cetacean on subtropical reefs in Brazil (Sazima et al. 2003). Parrotfishes and surgeonfishes may have more varied diets and nutritional ecology than previously believed, playing important roles as detritivores and microphages. However, there has been surprisingly little research documenting the role of coprophagy in the foraging dynamics of these fishes. Coprophagy has clear documentation and significance in terrestrial ecosystems (e.g., Soave and Brand 1991; Hirakawa 2001; Nalepa et al. 2001). It may also play an important role in nutrient transfer in marine ecosystems, such as providing a source of iron in the mesopelagic (Le Mézo and Galbraith 2021), nitrogen and phosphorus in temperate littoral systems (Pinnegar and Polunin 2006), carbon and nitrogen for marine invertebrates (Frankenberg and Smith 1967), and as a possible source of detrital organic matter for marine zooplankton recycling (Turner and Ferrante 1979). However, there is little documentation of the nutritional significance and transfer of energy through coprophagy on tropical coral reefs (Bailey and Robertson 1982; Robertson 1982).

Coral reefs are typically nutrient-limited ecosystems. Algae are generally considered to be of low-nutritional quality in the diets of herbivorous fishes (Horn 1989; Harmelin-Vivien 2002; Lobato et al. 2014; Shantz et al. 2017) as they are relatively low in protein and contain carbohydrates that are difficult to digest for fishes with a limited capacity for hindgut fermentation, such as parrotfishes (Crossman et al. 2005; Clements et al. 2016). Moreover, due to the relatively low abundance of nitrogen and phosphorus in algae, the growth of herbivorous fishes can be limited by nitrogen and phosphorus rather than energy (Hood et al. 2005; Benstead et al. 2014; Shantz et al. 2017; Schiettekatte et al. 2020). Fish feces may be a rich source of some of these nutrients and the nutritional value of fish feces can be equal to, or higher than, that of non-fecal food sources (Bailey and Robertson 1982). Coprophagous fishes often graze upon the feces of planktivores and higher trophic level fishes more frequently than fishes at lower trophic levels (Bailey and Robertson 1982; Robertson 1982). Planktivorous fish feces may be a particularly rich nutritional food source as they can be high in amino acids (Crossman et al. 2005), and their protein and lipid content may equal to or exceed that of algae, which dominates the diet of parrotfishes and surgeonfishes (Bailey and Robertson 1982). Therefore, fish fecal material may play an important role in the nutritional ecology of herbivorous reef fishes and nutrient cycling on coral reefs (Robertson 1982; Meyer and Schultz 1985; Smriga et al. 2010).

While recent studies have highlighted the importance of fish waste products on nutrient and energy cycling in coral reef food webs (Allgeier et al. 2017; Gil and Hein 2017; Ezzat et al. 2019), the role of coprophagy as a nutritional resource for fishes on coral reefs has received little attention (Bailey and Robertson 1982; Robertson 1982). Coprophagy may be an additional nutrient cycling mechanism to consider when assessing dietary preferences and nutritional needs of these key herbivores on coral reefs.

In this study, we investigated species-specific patterns of coprophagy of planktivorous Brown Chromis (*Chromis multilineata*) feces by parrotfishes and surgeonfishes on Caribbean reefs. Additionally, we analyzed the nutritional value of *C. multilineata* feces compared to other food sources targeted by parrotfishes and surgeonfishes. Our objectives were to: (1) quantify species-specific consumption of *C. multilineata* fecal pellets relative to total bites across food sources by parrotfishes and surgeonfishes, (2) determine the frequency at which these fecal pellets are consumed by parrotfishes, surgeonfishes, and other reef fishes, and (3) compare the nutritional value of these fecal pellets (in terms of proteins, lipids, carbohydrates, total calories, and micronutrients) to known values from other food sources. To our knowledge, this is the first study to

document coprophagy by herbivorous fishes in the Caribbean. This behavioral research, paired with the nutritional value of these ingested fecal pellets, advances our understanding of the grazing dynamics of nominally herbivorous fishes and the importance of nutrient cycling via the fish feces on coral reefs.

## Material and methods

### Study sites and species

We conducted this study from June to September 2019 on the Southern Caribbean island of Bonaire (Fig. S1). We surveyed four primary sites on the island's leeward coast (Karpata, Tolo, Cliff, and Bachelor's Beach), as well as two additional sites (Andrea I and Sweet Dreams) where we collected fish behavioral data for less abundant parrotfish species (described below). These sites were characterized by continuous fringing nearshore reefs. We conducted behavioral surveys of all three Caribbean surgeonfishes (*Acanthurus chirurgus*, *A. coeruleus*, and *A. tractus*—previously known as *A. bahianus*) and most reef-dwelling parrotfishes (*Scarus coeruleus*, *S. guacamaia*, *S. iseri*, *S. taeniopterus*, *S. vetula*, *Sparisoma aurofrenatum*, *S. chrysopterygum*, *S. rubripinne*, and *S. viride*). The three largest Caribbean parrotfishes—*S. coeruleus*, *S. guacamaia*, and *S. coelestinus*—were rare across these study sites. Therefore, we conducted additional behavioral surveys of *S. guacamaia* and *S. coeruleus* at the sites Andrea I and Sweet Dreams to obtain at least 6 behavioral surveys per species. *Scarus coelestinus* were not included in this study since we did not encounter this species during any surveys.

Additionally, we monitored the defecated fecal pellets of *C. multilineata*—an abundant planktivore on Caribbean reefs—to track the frequency at which these fecal pellets were consumed by fishes. *Chromis multilineata* often forage high in the water column to feed on plankton (Hobson 1973). As a result, their fecal pellets fall through the water column before reaching the benthos (see supplementary video footage), providing opportunities for coprophagous fishes to target them (Bray et al. 1986). We chose to focus on the fecal pellets of *C. multilineata* based on behavioral observations of parrotfishes and surgeonfishes. We observed that one individual *S. taeniopterus* and one *S. vetula* each consumed one fecal pellet of a Sergeant Major damselfish (*Abudefduf saxatilis*) compared to 876 total instances of consumption of *C. multilineata* fecal pellets by parrotfishes and surgeonfishes. These findings suggest that although they may occasionally consume fecal pellets of other fish species, parrotfishes and surgeonfishes on Bonaire predominantly target *C. multilineata* feces.

## Fish density surveys

To quantify the density of parrotfishes, surgeonfishes, and *C. multilineata* at each site, we used a roving diver method similar to Adam et al. (2015). We conducted timed swims while towing a Garmin GPS 72 receiver on a float to measure the distance traveled. We surveyed parrotfishes, surgeonfishes, and *C. multilineata* separately to ensure accurate counts of each taxa. During surveys, we recorded the species and fork length of all *C. multilineata*  $\geq 3$  cm, surgeonfishes  $\geq 10$  cm, and parrotfishes  $\geq 15$  cm within a 5 m wide belt (i.e., 2.5 m to either side of the observer). Some parrotfish species were less common at our study sites, while surgeonfishes and *C. multilineata* were more abundant. Therefore, we conducted 10 min surveys of surgeonfishes and *C. multilineata*, but 20–25 min surveys of parrotfishes to sample a sufficiently large area to accurately estimate the density of less common parrotfish species. Given the high abundance of *C. multilineata*, it was not feasible to record the exact length of every individual during the 10 min surveys. Therefore, we recorded the abundance of *C. multilineata* in bins of 3 to  $< 6$  cm, 6 to  $< 8$  cm, 8 to  $< 10$  cm, and 10 to  $< 12$  cm. We conducted four surveys of fish density per taxa per site across a range of depths from 4 to 18 m. We calculated the distance traveled between GPS points throughout the survey using the R package ‘geosphere’ (Hijmans 2019), then multiplied the survey distance by 5 m (the survey width) to calculate the total survey area.

## Fecal pellet observations

To determine the proportion of *C. multilineata* fecal pellets that were consumed by different coprophagous fishes, we monitored fecal pellets using a similar protocol to that of Robertson (1982). We followed haphazardly selected *C. multilineata* individuals until they defecated between 9:00 and 16:00 ( $n = 135$ ). Upon defecation, we recorded the depth of the fecal pellet in the water column (m) and the depth of the benthos (m) to calculate the distance above the benthos at which the individual defecated. Additionally, we calculated the rate at which fecal pellets sank ( $\text{m s}^{-1}$ ) by recording the time in seconds it took the fecal pellet to travel several meters or until it was consumed by a coprophage. We monitored each fecal pellet until it was consumed or reached the benthos and recorded whether the fecal pellet was inspected (approached but not consumed), tasted (ingested, then expelled), eaten, or not consumed. If the fecal pellet was inspected, tasted, or eaten, we recorded the species of the coprophagous fish (or genus in the case of a few difficult to distinguish *Stegastes* spp.). To minimize the potential effect of observers on fish behavior, we

maintained at least a 2 m distance from focal fishes and fecal pellets.

## Behavioral observations

To characterize the foraging patterns of parrotfishes and surgeonfishes, we conducted behavioral observations across study sites following Adam et al. (2015). We surveyed surgeonfishes  $\geq 10$  cm fork length and parrotfishes  $\geq 15$  cm, with the exception of the parrotfish *S. iseri*, for which we included individuals  $\geq 10$  cm due to their smaller length at sex change (Allsop and West 2003). Because parrotfishes are protogynous hermaphrodites and their diet may vary between initial and terminal phase individuals, we surveyed 6 initial phase and 6 terminal phase individuals of each species whenever possible. However, some parrotfish species and/or phases were rare across our study sites, therefore we surveyed a minimum of at least 6 individuals per species regardless of phase (see Table S1 for sample sizes). For surgeonfishes, we conducted 6 surveys per species. We conducted surveys between 9:00 and 16:00, a time frame during which parrotfishes and surgeonfishes are typically actively foraging and that is similar to that of previous studies of the foraging behavior of these fishes in the Western Atlantic (Bruggemann et al. 1994a; van Rooij et al. 1996; Bonaldo et al. 2006; Adam et al. 2015; Duran et al. 2019). Between 9:00 and 16:00, there appeared to be little relationship between the time of day on coprophagy rates (Fig. S2). We haphazardly selected focal fishes and allowed them approximately 2 min to acclimate to the diver prior to the survey. Thereafter, we followed the individual for 20 min and recorded the number of bites taken on feces and various non-fecal food sources. In a few rare instances, focal fishes swam out of sight prior to a full 20 min survey duration; we only included these data if the survey had lasted at least 15 min. To determine the defecation rates of *C. multilineata*, we haphazardly selected individuals  $\geq 5$  cm and observed them for 3 min ( $n = 50$ ). We recorded the fork length of each fish to the nearest centimeter and total number of defecations during the survey period.

## Fecal sample collection

To analyze the nutritional quality of *C. multilineata* feces, we collected samples of fecal pellets. To do so, we swam in the water column following haphazardly selected *C. multilineata* individuals and opportunistically collected fecal samples in 2.0 mL sterilized vials. To avoid diver contamination and ensure the collection of intact fecal pellets, we positioned the vials below feces and allowed them to sink directly into the tube. We collected three fecal pellets per vial ( $n = 50$  vials). We transported samples on ice to

our field research facility, then immediately froze samples at  $-18\text{ }^{\circ}\text{C}$ . We used an Air Sea Containers Bio-Freezer Pack to transport frozen samples from Bonaire to our laboratory facility at California Polytechnic State University, San Luis Obispo and kept samples frozen until we conducted fecal nutritional analyses.

### Fecal nutritional analyses

#### *Sample preparation*

The *C. multilineata* fecal samples we collected originally contained three fecal pellets per vial ( $n = 50$  vials); however, since individual pellets were small, we combined vials to yield enough material for nutritional analyses. To do so, we added 50 mL of deionized water to each vial for fecal homogenization and extraction and then combined the contents of five vials into sterile 50 mL Falcon centrifuge tubes (resulting in  $n = 10$  sample vials, each containing 15 fecal pellets). We homogenized samples using a probe sonicator and centrifuged them at 4700 xg for 60 s, then pipetted and discarded excess water from the surface. We then flash froze wet samples in liquid nitrogen immediately before placing them in a Labconco lyophilizer at  $-50\text{ }^{\circ}\text{C}$  with a 20 mTorr vacuum for 24 h. After freeze-drying samples, we recorded the dry weight of samples to the nearest 0.0001 g.

#### *Protein analysis*

To determine feces protein content, we conducted a bicinchoninic acid (BCA) assay. This method uses the reduction of  $\text{Cu}^{+2}$  to  $\text{Cu}^{+1}$  by protein to produce a colorimetric reaction and can be more tolerant to interfering substances than the Lowry method (Smith et al. 1985). We first placed 20 mg aliquots of homogenized, dried fecal material from each sample vial in 1.5 mL Eppendorf tubes and added 0.6 mL of 1 M sodium hydroxide (NaOH). We homogenized samples by securing tubes horizontally and vortexing them at room temperature using an orbital shaker at medium speed for 12 h (Montgomery and Gerking 1980; Crossman et al. 2005). We centrifuged samples at 12,000 xg for 5 min and transferred 300  $\mu\text{L}$  of supernatant from each sample into a new Eppendorf tube. We analyzed protein content using a Thermo Scientific™ Pierce™ Rapid Gold BCA Protein Assay Kit. We prepared and added kit working reagents to the BCA standard and samples in a flat-bottom 96-well plate and read absorbances in a spectrophotometer at OD 480 nm, then determined protein concentrations using the standard curve.

#### *Lipid analysis*

To determine feces lipid content, we conducted a colorimetric assay using the sulfo-phospho-vanillin (SPV) method (Chabrol and Charonnet 1937). This method is well-established for quantifying lipids and has been used in studies of marine fishes and invertebrates (Lu et al. 2008; Park et al. 2016). We extracted lipid content using an Abcam Chloroform-Free Lipid Extraction Kit. We placed 40–50 mg aliquots of fecal material in Eppendorf tubes ( $n = 10$ ) and added 500  $\mu\text{L}$  of the extraction buffer, then homogenized samples with a probe sonicator for 1–2 min. We kept samples on ice before and after sonication, then centrifuged them at 10,000 xg for 5 min at  $4\text{ }^{\circ}\text{C}$ . We collected the supernatant in microcentrifuge tubes and placed samples on an orbital shaker at room temperature for 15 min, then centrifuged them again at 10,000 xg for 5 min at room temperature. We collected the resulting supernatant and flash froze samples using liquid nitrogen. We resuspended the lipid extract in 50  $\mu\text{L}$  of the kit's suspension buffer and agitated it for 20 min at room temperature in a bath sonicator. We then determined lipid content using an Abcam Lipid Assay Kit. We prepared standards using dimethyl sulfoxide (DMSO) and placed standards and samples in a 96-well plate with sulfuric acid before heating them for 10 min to prime the lipid. We added vanillin that reacted colorimetrically with the lipid in the acidic solution, read absorbances in a spectrophotometer at OD 540 nm, and determined lipid concentrations using the standard curve.

#### *Ash content analysis*

To determine feces ash content, we transferred 150 mg ( $\pm 5$  mg) aliquots of wet sample into pre-labeled aluminum weigh boats ( $n = 10$ ). We dried samples in a convection oven at  $90\text{ }^{\circ}\text{C}$  until samples reached a constant weight (3 h) and then immediately recorded the sample dry weight to the nearest 0.0001 g. We placed the dried samples in a muffle furnace for 5 h at  $450\text{ }^{\circ}\text{C}$ , similar to McDermid and Stuercke (2003). We let samples cool for 5 h to room temperature, then immediately re-weighed samples to the nearest 0.0001 g to determine ash content.

#### *Carbohydrate and total caloric value calculations*

To determine feces carbohydrate content, we calculated carbohydrates by difference as follows: carbohydrates =  $100 - \text{proteins} - \text{lipids} - \text{ash}$ , where all variables are in % dry weight. This method is commonly used to determine total carbohydrates in food, including in fish feed nutrition studies (Southgate 1969; Opstvedt et al. 2003). Using the sample weight and percentage of proteins,



lipids, and carbohydrates, we estimated the caloric values of fecal pellets using established conversion factors based on those broadly accepted by the USDA: carbohydrate, 4.0 kcal g<sup>-1</sup>; lipid, 9.0 kcal g<sup>-1</sup>; protein, 4.0 kcal g<sup>-1</sup> (Atwater and Woods 1896; Maclean et al. 2003).

#### Micronutrient analysis

To quantify levels of several biologically important minor elements (calcium, magnesium, and phosphorus) and trace elements (copper, iron, and zinc) in feces, we sent 15 mg aliquots of homogenized, freeze-dried fecal material ( $n = 10$ ) to the Michigan Elemental Analysis Lab at the University of Michigan for analysis using an inductively coupled plasma mass spectrometer (Thermo Scientific iCAP Q ICP-MS). They digested the samples in 1 mL conc. Optima™ nitric acid (HNO<sub>3</sub>) using a CEM Mars 6 Microwave. Afterward, they diluted samples to 2% HNO<sub>3</sub> (v v<sup>-1</sup>) and analyzed them using the kinetic energy discrimination (KED) mode with helium gas to overcome polyatomic interferences. The data were standardized based on sample weight and reported in percent dry weight or parts per million (ppm), depending on the element.

#### Nutritional value of other food sources

To compare the nutritional value of *C. multilineata* fecal pellets to the other food sources commonly targeted by parrotfishes and surgeonfishes, we conducted a literature search for published nutritional values for brown, green, and red algae, the epilithic and endolithic algae matrix (EAM), and cyanobacteria. We limited this search to articles examining wild, tropical algae of genera that occur in the Caribbean region, as well as studies of the EAM and cyanobacteria sampled from tropical regions (see Supplementary Material for a list of all articles). We compiled the values of total carbohydrates, lipids, and proteins (% dry weight) and the total caloric value (kcal g<sup>-1</sup>) (see Table S2 for sample sizes). In addition, we compiled published values on the content of magnesium, calcium, and phosphorus (% dry weight) and copper, iron, and zinc (parts per million) for brown, green, and red algae (see Table S3 for sample sizes). We were unable to find published values on the content of these micronutrients for the tropical EAM and cyanobacteria.

#### Statistical analysis

We conducted statistical analyses using R v3.6.2 and the ‘tidyverse’ package suite (R Core Team 2019; Wickham et al. 2019; RStudio Team 2020). We conducted a correlation test to analyze the relationship between defecation rate and fork length (cm) of *C. multilineata*. We conducted

a binomial logistic regression analysis to examine if there were significant differences in the proportion of the total number of bites by fishes on *C. multilineata* feces in response to the distance above the benthos at which the *C. multilineata* defecated ( $m$ ), the rate at which the feces sank ( $m\ s^{-1}$ ), and the interaction between these variables.

To evaluate the difference in the proportion of fecal pellets consumed by different parrotfish and surgeonfish species after accounting for differences in the proportional abundance of these fishes, we used a simulation-based test. We generated 1000 random samples of the proportion of feces consumed by each fish species from simulation based on their proportional abundance. We compared the distribution of possible proportions of coprophagy from the simulated data to the actual observed proportions. For the test, we restricted our comparisons to parrotfishes and surgeonfishes for which we conducted density surveys, not other fish taxa. However, observations of coprophagy by other fish taxa were rare. Of the 135 fecal pellets we monitored, a total of 104 were consumed by parrotfishes and surgeonfishes and only 10 were consumed by other fish taxa.

To analyze species-specific differences in coprophagous behavior, we used Kruskal–Wallis tests to compare: (1) the percentage of bites on fecal pellets relative to total bites and (2) the bite rate on fecal pellets (bites per minute) by parrotfish and surgeonfish species. To determine which species differed significantly, we conducted post hoc comparisons using Dunn’s tests. For these analyses, we used the ‘stats’ and ‘dunn.test’ R packages (Dinno 2017; R Core Team 2019).

## Results

### Species-specific rates of coprophagy

*Chromis multilineata* were abundant across our study sites, with an average density that ranged from 1.6 ( $\pm 0.3$  SE) to 3.1 ( $\pm 1.1$  SE) individuals per m<sup>2</sup>. Based on observations, we estimated that *C. multilineata* had an average defecation rate of 5.6 ( $\pm 1.3$  SE) fecal pellets per hour ( $n = 50$  behavioral surveys). We found no relationship between *C. multilineata* defecation rate and fork length (correlation test,  $p$  value  $> 0.05$ ), but the range of fish fork lengths for individuals we observed was small (6–8 cm).

We followed 135 *C. multilineata* fecal pellets and observed that 84.4% were consumed, 3.7% were tasted but not ingested, 0.7% were inspected, and 11.1% were not inspected or consumed prior to reaching the benthos. Of the 114 fecal pellets consumed by fishes, 91.2% were consumed by parrotfishes or surgeonfishes alone, while 8.8% were consumed by other fishes (*Abudefduf saxatilis*,

*Pomacanthus paru*, *Stegastes partitus*, *S. planifrons*, and other *Stegastes* spp.). The parrotfish *S. taeniopterus* and surgeonfish *A. coeruleus* were more frequent coprophages, where of the total 114 observed instances of coprophagy of *C. multilineata* fecal pellets, 44.7% were by *S. taeniopterus* and 26.3% were by *A. coeruleus*. We observed that coprophagous fishes regularly swam high in the water column to consume descending fecal pellets (see supplementary video footage). The range of depths at which we observed feces being defecated by *C. multilineata* spanned from approximately 3 to 14 m above the benthos, while the range at which we observed feces being consumed spanned from approximately 5 to 18 m above the benthos. Based on our observations, we estimate that fecal pellets sank at an average rate of  $0.02 \text{ m s}^{-1}$  ( $\pm 0.001$  SE). We found no evidence of a significant difference in the likelihood of feces being consumed based on the distance above the benthos at which *C. multilineata* defecated (m), rate that feces sank ( $\text{m s}^{-1}$ ), or the interaction between these variables ( $p$  value  $> 0.05$ ).

Based on observations of *C. multilineata* fecal pellets, as well as surveys of the foraging behavior of parrotfishes and surgeonfishes, we found that all three surgeonfishes (*Acanthurus chirurgus*, *A. coeruleus*, and *A. tractus*) and many parrotfish species surveyed (*Scarus coeruleus*, *S. iseri*, *S. taeniopterus*, *S. vetula*, *Sparisoma aurofrenatum*, and *S. viride*) consumed *C. multilineata* feces (Fig. S3). For some species, coprophagy was rare; we observed *S. coelestinus* consuming fecal pellets during surveys of their foraging behavior but not during our fecal pellet observations, and we only observed *S. vetula* engaging in coprophagy during fecal pellet observations but not during foraging behavioral surveys. Several parrotfish species were not observed engaging in coprophagy using either survey method (*S. guacamaia*, *S. chrysopterus*, and *S. rubripinne*).

#### *Analysis of coprophagy based on fecal pellet observations*

There was a significant difference in the observed proportion of feces consumed by different parrotfish and surgeonfish species than predicted from the distribution of possible simulated proportions based on species proportional abundance (Fig. 1). Our findings suggest that *S. taeniopterus* and *A. coeruleus* consumed a greater proportion of feces (Simulation-based test,  $p$  value  $< 0.001$  and  $0.031$ , respectively) and that *S. coeruleus*, *S. guacamaia*, *S. chrysopterus*, *S. rubripinne*, and *S. viride* consumed a lesser proportion of feces than predicted after accounting for the relative density of parrotfish and surgeonfish species ( $p$  value  $< 0.001$  for all, Fig. S4).

#### *Analysis of coprophagy based on fish behavioral observations*

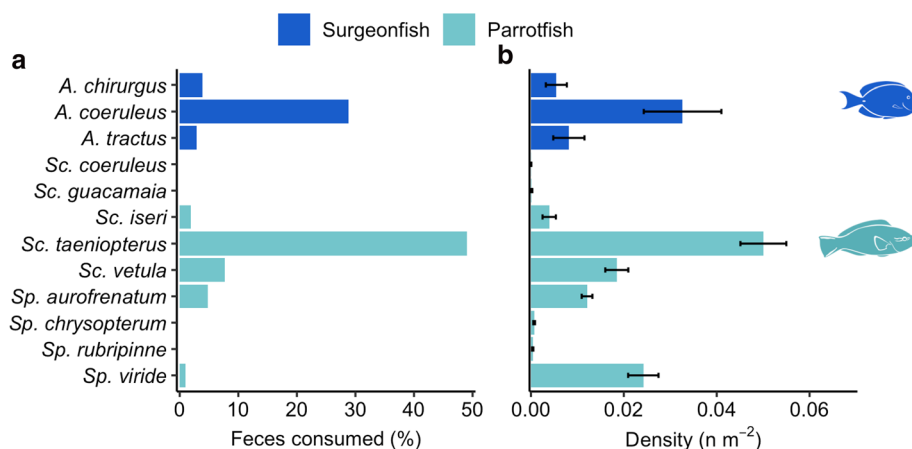
Based on surveys of the foraging behavior of parrotfishes and surgeonfishes (Fig. 2), there was a significant difference in the median percentage of bites on fecal pellets compared to total bites across food sources among species (Kruskal–Wallis test,  $X^2 = 58.313$ ,  $df = 11$ ,  $p$  value  $< 0.001$ ). Our results suggest that *A. coeruleus*, *S. iseri*, *S. taeniopterus*, and *S. aurofrenatum* had a significantly higher percentage of bites on feces compared to *S. guacamaia*, *S. vetula*, and *S. chrysopterus* (Dunn's test, adj.  $p$  value  $< 0.05$  for all; Table S4). Additionally, *S. taeniopterus* had a significantly higher percentage of bites on feces compared to *S. viride* (Dunn's test, adj.  $p$  value =  $0.044$ ). There was also a significant difference in the median bite rate (bites per minute) on fecal pellets among species (Kruskal–Wallis test,  $X^2 = 58.285$ ,  $df = 11$ ,  $p$  value  $< 0.001$ ). Our findings suggest that *A. coeruleus*, *S. iseri*, and *S. taeniopterus* had significantly higher bite rates on fecal pellets compared to *S. guacamaia*, *S. vetula*, *S. chrysopterus*, and *S. viride* (Dunn's test, adj.  $p$  value  $< 0.05$  for all; Table S5).

#### **Nutritional value of feces**

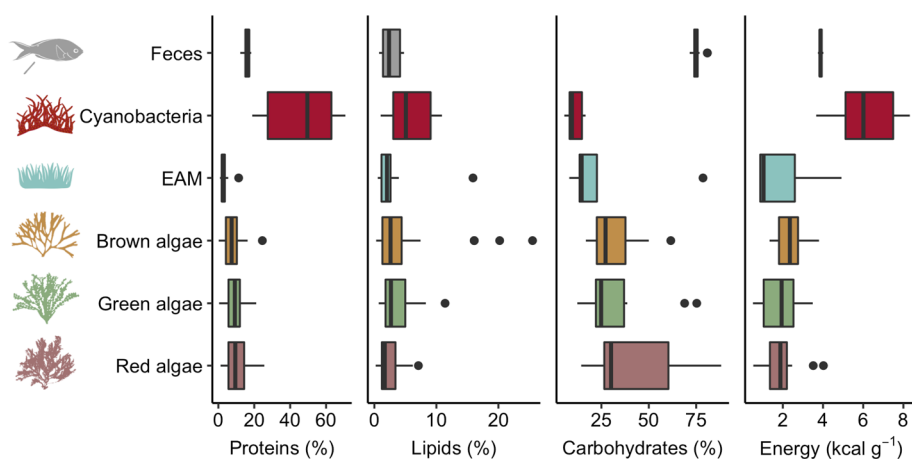
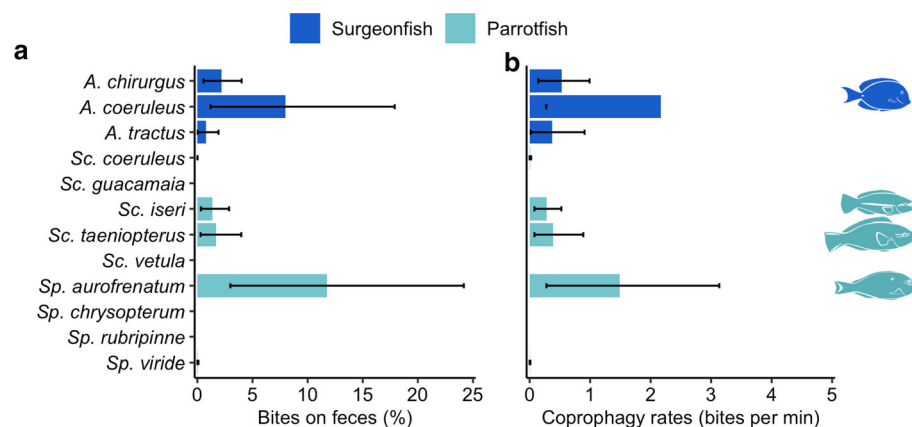
We estimated that the *C. multilineata* fecal pellets contained a median protein content of 16.6% dry weight. This was greater than the median of reported values for brown, green, and red algae, as well as the EAM, though lower than that of cyanobacteria (Fig. 3; Table S6). We estimated that the median lipid content of fecal pellets was 2.4% dry weight, which was similar to the median of reported values for brown, green, and red algae, as well as the EAM, but lower than that of cyanobacteria. Based on calculations of percent carbohydrates in feces, we estimated the median carbohydrate content in fecal pellets to be 75.1% dry weight, which was far higher than values of all other food items considered. Based on calculations of fecal pellet caloric value, we estimated that they had a median of  $3.9 \text{ kcal g}^{-1}$ , which was higher than the median for brown, green, and red algae ( $2.4$ ,  $1.9$ , and  $1.9 \text{ kcal g}^{-1}$ , respectively) and the EAM ( $1.0 \text{ kcal g}^{-1}$ ), but lower than that of cyanobacteria ( $6.0 \text{ kcal g}^{-1}$ ).

In addition to these macronutrients, coprophagous fishes may also derive nutritional value from minor and trace elements in fish feces. We found that concentrations of several minor elements (calcium, magnesium, and phosphorus) and trace elements (copper, iron, and zinc) in the *C. multilineata* fecal pellets were well above the median values reported for a range of tropical algal species (Fig. 4; Table S7). The median levels of calcium, copper, magnesium, phosphorus, and zinc were higher in fecal pellets

**Fig. 1** **a** Percentage of *Chromis multilineata* fecal pellets consumed by parrotfish and surgeonfish species ( $n = 104$  total fecal pellets consumed by parrotfishes and surgeonfishes). **b** Mean density ( $n\ m^{-2}$ ) of herbivorous parrotfishes and surgeonfishes across sites  $\pm$  SE ( $n = 4$  sites)



**Fig. 2** **a** Median percentage of bites by parrotfishes and surgeonfishes on *Chromis multilineata* fecal pellets compared to other food sources. **b** Median coprophagy rates (bites per min) by parrotfishes and surgeonfishes. We used bootstrapping to generate 95% confidence intervals based on 1000 sampling iterations ( $n = 6$ – $12$  20 min behavioral observations per species)



**Fig. 3** Macronutrient levels (% dry weight) and total caloric values of *Chromis multilineata* fecal pellets compared to values from the literature of other food sources targeted by parrotfishes and surgeonfishes (see Table S2 for sample sizes). EAM values include the epilithic algae matrix, endolithic algae matrix, turf algae, and

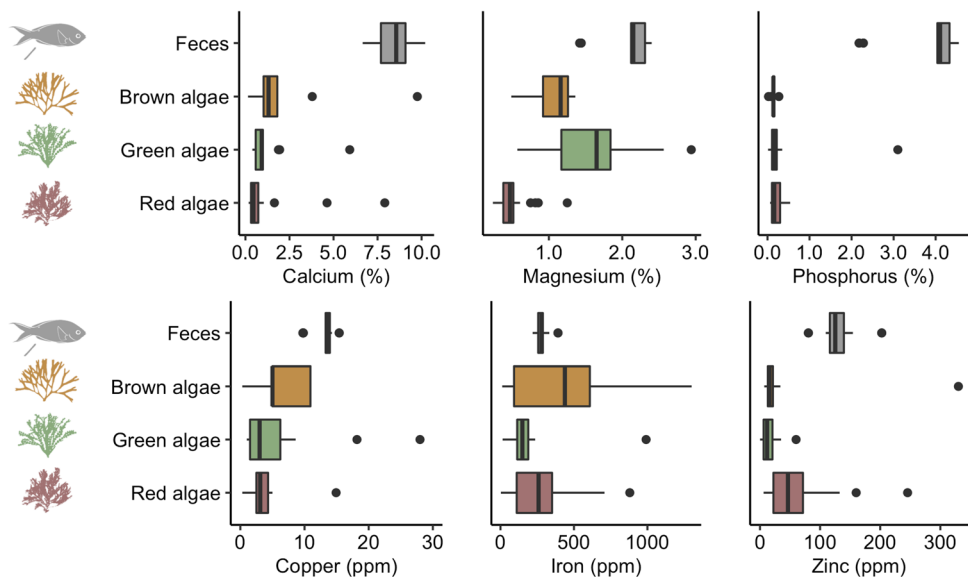
associated detritus. The boxplots show the median and interquartile range (IQR), with whiskers extending to values up to 1.5 times the IQR and outlying data displayed as individual points. Ash content values are presented in Table S6

compared to the median of reported values for tropical brown, green, and red algae. Median concentrations of iron in fecal pellets were also greater than values for green

algae, though similar to that of red algae and less than that of brown algae.



**Fig. 4** Minor elements (% dry weight) and trace elements (parts per million) in *Chromis multilineata* fecal pellets compared to values from the literature of other food sources targeted by parrotfishes and surgeonfishes (see Table S3 for sample sizes). The boxplots show the median and interquartile range (IQR), with whiskers extending to values up to 1.5 times the IQR and outlying data displayed as individual points



## Discussion

This study provides the first documentation and quantitative analyses of coprophagous behavior of nominally herbivorous fishes in the Caribbean, as well as early insights into the potential nutritional values of this behavior. We found that consumption of fish feces was prevalent among Caribbean parrotfishes and surgeonfishes, where six of the nine parrotfish species surveyed and all three surgeonfish species actively consumed the feces of an abundant planktivore, *Chromis multilineata*. While other species were observed consuming *C. multilineata* feces, we found that over 90% of ingested fecal pellets were consumed by parrotfishes and surgeonfishes alone. Similar to our results, a previous study that found that over 90% of fecal pellets from zooplanktivorous fishes were consumed by reef fishes in the Indo-Pacific and nearly 97% of ingested fecal pellets were consumed by herbivores and detritivores alone (Robertson 1982). Furthermore, we observed that several species regularly swam high in the water column to forage on descending fecal pellets as they slowly sank. Since parrotfishes and surgeonfishes typically graze on EAM, some macroalgae, and other food items on the reef benthos (Adam et al. 2015; Dromard et al. 2015; Duran et al. 2019), this foraging behavior strongly suggests that these species are actively targeting these feces.

Such high consumption rates, broad utilization, and active targeting of fecal pellets across a range of species suggest that these fishes are deriving some nutritional benefit beyond what they obtain through their nominally herbivorous diets. We found that these *C. multilineata* fecal pellets were rich in macronutrients such as protein and some potentially limiting micronutrients, similar to findings from previous studies of *Chromis* spp. fecal pellets in

other tropical and temperate reefs (Bailey and Robertson 1982; Geesey et al. 1984; Pinnegar and Polunin 2006). While there are species-specific differences in levels of coprophagy, these planktivore feces were targeted by a wide variety of parrotfish and surgeonfish species on Bonaire. Our observations of high levels of coprophagy of planktivore feces by herbivorous fishes in the Caribbean are similar to those of a previous study in the Indo-Pacific (Robertson 1982), suggesting that these feces may be an important source of nutrition for herbivorous fishes in other regions.

Parrotfishes and surgeonfishes have long been recognized for their important functional role in grazing upon and removing turf algae and macroalgae from coral substrate and rubble, a behavior which is believed to mediate competition between coral and algae. However, recent work suggests that these herbivores vary significantly in their foraging behavior interspecifically, ontogenetically, and as a function of resource availability and habitat type (Adam et al. 2015; Smith et al. 2018; Duran et al. 2019). The prevailing hypothesis has been that parrotfishes and surgeonfishes primarily target algae. Yet, recent studies suggest that these nominal herbivores may target detritus and protein-rich microscopic autotrophs—such as cyanobacteria—within or upon the algal-covered substrate or epiphytic on algae (Crossman et al. 2001; Clements et al. 2016; Mendes et al. 2018; Cissell et al. 2019; Nicholson and Clements 2020), as well as other food items such as corals and sponges (Burkepile et al. 2019; Rempel et al. 2020). Our findings suggest that fecal pellets may be another important but overlooked component in the diets of these fishes.

A previous study by Robertson (1982) found that coprophagy is widespread among herbivorous and

detritivorous coral reef fishes in the Indo-Pacific and suggested that ‘high-quality’ feces may be a more nutritious energy source than algae. Moreover, that study found that the feces of zooplanktivorous fishes—including a few *Chromis* spp.—were more frequently consumed than those of corallivores and herbivores. Interestingly, based on behavioral surveys, we found that 99.8% of the fecal pellets consumed by parrotfishes and surgeonfishes were from a single planktivorous species, *C. multilineata*. Since our behavioral data strongly suggest that herbivores on Bonaire were targeting the fecal pellets of this planktivore, we examined their nutritional content and compared our results to published values for brown, green, and red algae, as well as the EAM and cyanobacteria. Our analyses of macronutrients reveal that these planktivore feces may have higher protein levels than those reported for all food types except cyanobacteria, higher total caloric energy than all algal food types, and higher levels of carbohydrates than all food sources examined. These findings are similar to those of Dethier and colleagues that suggested that feces of a marine invertebrate may have higher protein content than algae (Dethier et al. 2019). Additionally, Bailey and Robertson (1982) found that the feces of the planktivore *Chromis atripectoralis* were rich in protein, lipids, and total calories compared to the feces of other coral reef fishes. The comparatively high protein levels and total caloric energy in fecal pellets we sampled suggest that *C. multilineata* feces, like cyanobacteria and the EAM, may be targeted to supplement the relatively lower-quality algae-based diets of parrotfishes and surgeonfishes.

While previous studies have examined the nutritional value of some of the food sources targeted by herbivores on coral reefs (Bruggemann et al. 1994a, b; Crossman et al. 2001), there are still knowledge gaps in the nutritional content of the EAM, cyanobacteria, and sessile invertebrates such as sponges and corals that these fishes are known to target. A recent review questioned whether algal resources are sufficient to support the high biomass of herbivorous fishes found on coral reefs (Clements et al. 2016). This work highlights the importance of integrating anatomical and biochemical analyses in addition to feeding observations to understand the dietary targets and nutritional ecology of nominally herbivorous fishes more fully.

Our findings suggest that planktivore fecal pellets are not only rich in macronutrients such as proteins and carbohydrates but may also be an important source of micronutrients. Coral reefs are commonly nutrient-poor ecosystems and may be limited in nutrients essential for metabolic processes (Howarth 1988; Watanabe et al. 1997). For example, the availability of some micronutrients, such as phosphorus and iron, may be limiting to the growth of reef organisms (Howarth 1988; Schiettekatte et al. 2020). Furthermore, elements such as copper and zinc

are critical for enzyme function and metabolic pathways and must be obtained in part from dietary intake (Hilton 1989; Watanabe et al. 1997). The feces of planktivores may be an important source of micronutrients such as nitrogen and phosphorus for herbivorous species that consume a nutrient-poor diet (Pinnegar and Polunin 2006). We found higher levels of calcium, copper, phosphorus, and zinc in *C. multilineata* feces compared to reported values for brown, green, and red macroalgae. The median phosphorus levels of these fecal pellets were over 25 times that of any algae taxa; this is notable as the growth of corals, reef algae, and other organisms may be limited by these nutrients (Howarth 1988). The high levels of phosphorus and calcium in fecal pellets may also drive the high levels of coprophagy we observed, since these elements cannot solely be taken up from the environment via gills and therefore are required from the diet (Sundell and Björnsson 1988; Hilton 1989; Flik and Verboost 1993). Furthermore, these micronutrients interact with one another in complex ways based on presence and quantity within a diet, therefore acting as important supplements (Hilton 1989). A previous study reported higher levels of phosphorus, copper, iron, and zinc in *C. multilineata* feces from St. Croix compared to values we observed from Bonaire (Geesey et al. 1984), suggesting that the feces of this planktivore may be a rich source of nutrients in other areas of the Caribbean. Broadly, these findings suggest that coprophagy may supply an important source of micronutrients, particularly for herbivorous fishes.

While this study provides the first documentation of coprophagy by herbivorous fishes in the Caribbean and insights into the nutritional benefits of this behavior, additional research is needed to investigate the importance of coprophagy in nutrient cycling on coral reefs. A recent study suggests that fish growth can be limited by available nitrogen and phosphorus, particularly for herbivores and detritivores (Schiettekatte et al. 2020). Fish excretion may provide an important supply of nutrients in marine ecosystems, including nitrogen and phosphorus (Meyer and Schultz 1985; Allgeier et al. 2014; Schiettekatte et al. 2020), but little work has explored the importance of fish egestion in these processes (Bailey and Robertson 1982; Geesey et al. 1984; Pinnegar and Polunin 2006). Our findings support that fish feces may play an undervalued role in nutrient cycling on coral reefs. Future work evaluating patterns of coprophagy and assimilation of these nutrients across fish ontogenetic stages will provide further insight into the importance of this behavior for fish nutrition and bioenergetics.

While coprophagy has been observed by coral reef fishes, including parrotfishes and surgeonfishes in the Indo-Pacific (Robertson 1982) and a surgeonfish in subtropical reefs in Brazil (Sazima et al. 2003), this process has not

been well-documented in the vast majority of coral reef ecosystems. As a result, we are uncertain if this behavior is unique to Bonaire—an island where *C. multilineata* are abundant—or occurs throughout other areas of the Caribbean. Future work should examine how common this behavior is in other coral reef ecosystems, as well as whether the degree of coprophagous behavior is dependent in part on the availability of high-quality fecal pellets, such as that of planktivorous fishes. Additionally, while we observed that most parrotfishes and all surgeonfishes engaged in coprophagy, *S. taeniopterus* and *A. coeruleus* had particularly high levels of coprophagy. Gut content and stable isotope analyses suggest that *A. coeruleus* may have a higher consumption of fleshy macroalgae, algal turf, detritus, and invertebrates compared to other surgeonfishes and parrotfishes (Dromard et al. 2015). In contrast, research suggests that *S. taeniopterus* may have more dietary overlap with parrotfishes such as *S. iseri*, *S. vetula*, and *S. viride*, consuming more fleshy macroalgae and coral in addition to algal turf and detritus (Dromard et al. 2015; Smith et al. 2018), as well as benthic cyanobacterial mats (Cissell et al. 2019). Future work examining the nutritional value of the various food sources targeted by parrotfish and surgeonfish species (e.g., specific carbohydrate, amino acid, and lipid profiles) and assimilation of nutrients may help to reveal why certain species are more coprophagous than others.

Our novel observations of coprophagy by nominally herbivorous fishes in the Caribbean suggest that fish feces may be an important, yet previously understudied food resource for parrotfishes and surgeonfishes. While there were species-specific differences in the proportional levels of coprophagy and rates of fecal pellet consumption, we observed that most parrotfish species and all three surgeonfish species engaged in this behavior to some degree. Additionally, our findings suggest that these fecal pellets may have higher nutritional values of proteins, total calories, and certain micronutrients compared to macroalgae and the EAM. This study highlights the potential nutritional benefits of coprophagy and improves our understanding of the foraging ecology of nominally herbivorous fishes.

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**Data availability** All data and scripts are available in a publicly accessible GitHub repository ([https://github.com/hannahrempel/coprophagy\\_2022](https://github.com/hannahrempel/coprophagy_2022)). Data are also available from the NOAA National Centers for Environmental Information (NCEI Accession 0,242,462) at <https://www.ncei.noaa.gov/archive/accession/0242462>.

#### Declarations

**Conflict of interest** On behalf of all authors, the corresponding author states that there is no conflict of interest.

**Ethical approval** This research was conducted under permits from the Openbaar Lichaam Bonaire (Archive No. 2018012755) and the Rijkswaterstaat Zee en Delta (RWS-2018/51358). On behalf of all authors, the corresponding author states that this research was conducted in compliance with ethical standards.

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